DOES ZINC OVERDOSE IN RAT DIET ALTER CU, FE, MN, AND ZN CONCENTRATIONS IN A TAPEWORM HOST?*

I. Jankovská¹, V. Sloup¹, J. Száková², J. Magdálek¹, B. Horáková¹, I. Langrová¹

 ¹Czech University of Life Sciences Prague, Faculty of Agrobiology, Food and Natural Resources, Department of Zoology and Fisheries, Prague, Czech Republic
²Czech University of Life Sciences Prague, Faculty of Agrobiology, Food and Natural Resources, Department of Agroenvironmental Chemistry and Plant Nutrition, Prague, Czech Republic

We evaluated Cu, Fe, Mn, and Zn concentrations in the bone, muscle, testes, intestine, liver, kidneys and tapeworm parasites *Hymenolepis diminuta* of rats from four groups: 12 animals given zinc lactate (120 mg/rat and week) in feed mixture (M0 group); six animals given zinc lactate (120 mg/rat and week) in feed mixture and infected with tapeworms (MT group); six control animals fed a standard mixture of ST-1 for rats (00 group); and six control animals fed a standard mixture of ST-1 for rats and infected with tapeworms (0T group). The experiment was conducted over a six-week period. In our study, tapeworm presence decreased element concentrations in the majority of rat tissues. Tapeworms accumulated higher levels of zinc and manganese than did the majority of host tissues; however, they accumulated very little iron and copper in comparison to the host tissues. Zinc overdosing increased manganese concentrations in rat tissues; zinc overdosing also seemed to protect the liver from absorption of Fe by tapeworms.

Hymenolepis diminuta, Rattus norvegicus, accumulation, zinc lactate, manganese, iron, copper



doi: 10.2478/sab-2018-0015 Received for publication on-July 19, 2017 Accepted for publication on November 22, 2017

INTRODUCTION

Risk element contamination of the environment is a global problem (G i l - J i m e n e z et al., 2017; K i m et al., 2017; V y m a z a l, 2017). Using a large number of manufactured products with a wide range of applications is becoming more common. This has resulted in the general population becoming increasingly exposed to a wide variety of xenobiotics that may cause adverse health effects (J i m e n e z - D i a z et al., 2016; K u l m a et al., 2017). Studies dealing with animal nutrition are still required due to increasing environmental contamination (B u r g e s et al., 2016; H e n r i q u e z - H e r n a n d e z et al., 2016; P a v l o v i c et al., 2016), which affects food quality. Among the widely discussed elements, zinc belongs to the most intensively investigated ones, because of the worldwide utilization of this element resulting in potential contamination of the environment (S t r a c h e l et al., 2016).

Zinc is an essential trace element necessary for normal human functioning. It serves as an enzyme cofactor and protects cell membranes from lysis caused by complement activation and toxin release (Sloup et

^{*} Supported by the Internal Grant Agency of the Czech University of Life Sciences Prague (CIGA), Project No. 20152021, and by the Grant Agency of the Czech Republic (GA CR), Project No. 13-18154S. The authors declare no potential conflicts of interest.

al., 2017). Zinc is not stored in the body; therefore, dietary intake is required. Zinc lactate is a zinc salt of lactic acid. This product aids in the digestion and metabolism of phosphorus, and is necessary for protein synthesis and blood stability. Zinc lactate is commonly used as a dietary supplement for both humans and animals (Sloup et al., 2016).

Parasitic diseases are very common in animals, especially in farm animals bred in high concentrations; parasitoses can be very dangerous (K y r i a n o v a et al., 2017). However, some parasites as intestinal helminths can be also beneficial to their hosts. It is known that cestodes and acanthocephalans can decrease heavy metal concentrations in a host body (S u r e s et al., 2002; S u r e s, 2004).

We worked with experimental animal models (laboratory rats: *Rattus norvegicus* var. *alba*) infected with rat tapeworms (*Hymenolepis diminuta*); rats were later given high doses of zinc lactate, which is commonly used by humans as a dietary supplement. The aim of this research was (*i*) to determine how tapeworms affect the accumulation of zinc, and (*ii*) to assess the potential shifts of other micronutrients (Cu, Fe, Mn) in a host given a zinc overdose.

Hypothesis: tapeworms decrease element concentrations in a host given Zn overdoses.

MATERIAL AND METHODS

This experiment was conducted over a six-week period with thirty male Wistar rats divided into the following groups: twelve animals given zinc lactate (120 mg/rat and week) in feed mixture (M0 group); six animals given zinc lactate (120 mg/rat and week) in feed mixture and infected with tapeworms (MT group); six control animals fed a standard mixture of ST-1 for rats (00 group); and six control animals fed a standard mixture of ST-1 for rats and infected with tapeworms (0T group). The experimental design and animals used in this study are described in detail in J a n k o v s k a et al. (2016) and S l o u p et al. (2016).

^{mg/kg} Zn 200 J **L**

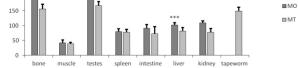


Fig.1a. Zinc concentrations (mg kg–1) in rat tissues and rat tapeworm M0 = rats with zinc lactate in feed mixture, MT = rats with zinc

Statistical analysis

Zn, Cu, Mn, and Fe concentrations and their statistical differences were compared within groups using the nonparametric Mann-Whitney U test. The differences were considered significant at P < 0.05. All computations were carried out using the STATISTICA software, Version 10 (Statsoft, USA).

RESULTS

In our study, tapeworm presence decreased element concentrations in the majority of rat tissues (Figs. 1a, b -4a, b). Tapeworms also accumulated higher levels of zinc and manganese than did the majority of host tissues (Fig. 1a, b and Fig. 3a, b). As expected, Zn overdosing significantly increased Zn concentrations in rat testes (Table 1a). In rats with tapeworm infection, zinc levels significantly increased in the testes, spleen, and in the tapeworm of rats with tapeworm infection; however, zinc overdosing surprisingly decreased Zn concentrations in the bone of rats with tapeworm infection (Table 1b).

DISCUSSION

The majority of elements found in humans and animals enter through the oral route and are subsequently absorbed in the digestive tract. This absorption process significantly interferes with gastrointestinal parasites. This is especially true for acanthocephalans and also tapeworms which receive nutrients through the tegument, a metabolically active body surface (S u r e s et al., 2000 a, b, 2002). Tapeworms are able to accumulate considerable amounts of metals, thereby reducing their concentrations in host tissues (J a n k o v s k a et al., 2010 a, b; C a d k o v a et al., 2013; B r o z o v a et al., 2015). Due to a close relationship between the local immune system and epithelial cells in the gastrointestinal tract, local immune reactions

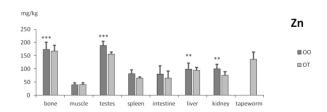


Fig. 1b. Zinc concentrations (mg kg-1) in rat tissues and rat tapeworm 00 = control rats, 0T = control rats with tapeworm infection statistically significant difference between groups ***P ≥ 0.001 , **P ≥ 0.05 , *P ≥ 0.01

Table 1a. Zn, Cu, Mn, and Fe concentrations (mg kg^{-1}) in rat tissues

| Zn | Bone | | Muscle | | Testes (*) | | Spleen | | Intestine | | Liver | | Kidney | | |
|------------------|------------|-------|-------------|-------|--------------|--------------|--------------|--------------|-----------|-----------------|-----------|--------|--------|--------------|--|
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | |
| 00 | 175.3 | 173.4 | 42.2 | 39.4 | 189.3 | 189.2 | 82.8 | 81.1 | 82.7 | 80.6 | 100.2 | 98.7 | 98.5 | 100.0 | |
| M0 | 193.6 | 193.2 | 43.9 | 41.9 | 197.6 | 199.8 | 77.8 | 79.6 | 90.7 | 91.5 | 104.2 | 102.5 | 103.5 | 110.0 | |
| Cu | bone | | muscle | | testes | | spleen (***) | | intestine | | liver | | kidney | | |
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | |
| 00 | 0.7 | 0.6 | 3.3 | 3.2 | 11.5 | 11.3 | 6.3 | 6.2 | 5.1 | 4.6 | 13.9 | 13.4 | 27.0 | 27.0 | |
| M0 | 1.0 | 0.7 | 3.1 | 3.0 | 11.6 | 11.6 | 4.1 | 4.1 | 5.2 | 5.0 | 14.4 | 14.4 | 30.5 | 29.8 | |
| Mn | bone (**) | | muscle (*) | | testes | testes (***) | | spleen (***) | | intestine (***) | | liver | | kidney (***) | |
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | |
| 00 | 0.3 | 0.3 | 0.3 | 0.2 | 1.7 | 1.7 | 0.8 | 0.8 | 2.7 | 2.4 | 7.3 | 7.5 | 3.0 | 3.1 | |
| M0 | 0.4 | 0.4 | 0.3 | 0.3 | 2.0 | 2.0 | 6.3 | 5.7 | 3.3 | 3.2 | 7.3 | 7.3 | 3.5 | 3.6 | |
| Fe | bone (***) | | muscle (**) | | testes (***) | | spleen | | intestine | | liver (*) | | kidney | | |
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | |
| 00 | 92.52 | 85.25 | 68.12 | 68.62 | 160.95 | 161.39 | 5187.45 | 4340.49 | 59.12 | 59.39 | 671.45 | 667.68 | 351.77 | 337.23 | |
| M0 | 53.04 | 54.20 | 40.40 | 37.14 | 132.81 | 132.62 | 5103.33 | 4543.09 | 50.07 | 50.20 | 508.85 | 517.50 | 376.46 | 375.18 | |

M0 = rats with zinc lactate in feed mixture, 00 = control rats, A = arithmetic mean, M = median*weak significance (P < 0.05), **medium sig-

nificance (P < 0.005), ***strong significance (P < 0.0005)

ll computations were done using program STATISTICA Version 10 (Statsoft, USA)

| Zn | Bone (*) | | Muscle | | Testes (*) | | Spleen (*) | | Intestine | | Liver | | Kidney | | Tapeworm (*) | |
|------------------|----------|-------|--------|-------|-------------|--------|------------|---------|-----------|-------|--------|--------|------------|--------|--------------|-------|
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | А | М |
| 0T | 167.0 | 168.4 | 41.2 | 41.2 | 155.0 | 156.7 | 68.4 | 65.6 | 73.2 | 65.3 | 89.0 | 94.9 | 83.0 | 75.9 | 136.3 | 136.6 |
| MT | 151.7 | 159.7 | 39.1 | 38.6 | 167.6 | 168.0 | 78.0 | 78.2 | 77.9 | 72.5 | 84.0 | 82.0 | 85.1 | 78.1 | 158.1 | 149.2 |
| Cu | bone | | muscle | | testes (*) | | spleen | | intestine | | liver | | kidney | | tapeworm | |
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | A | М |
| 0T | 0.9 | 0.7 | 3.1 | 3.1 | 11.1 | 11.1 | 5.4 | 4.6 | 5.1 | 5.0 | 12.0 | 12.5 | 27.1 | 27.4 | 5.9 | 5.1 |
| MT | 0.7 | 0.7 | 3.2 | 3.1 | 10.4 | 10.5 | 4.3 | 4.2 | 4.8 | 5.0 | 12.0 | 12.2 | 28.1 | 29.2 | 5.6 | 5.3 |
| Mn | bone | | muscle | | testes (**) | | spleen | | intestine | | liver | | kidney (*) | | tapeworm | |
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | А | М |
| 0T | 0.4 | 0.3 | 0.2 | 0.3 | 1.5 | 1.5 | 0.7 | 0.7 | 3.8 | 3.9 | 5.5 | 5.4 | 2.6 | 2.4 | 3.4 | 3.3 |
| MT | 0.3 | 0.3 | 0.3 | 0.3 | 1.4 | 1.4 | 0.7 | 0.7 | 4.8 | 4.4 | 5.4 | 5.3 | 2.3 | 2.3 | 3.6 | 3.8 |
| Fe | bone | | muscle | | testes | | spleen | | intestine | | liver | | kidney | | tapeworm | |
| Group/ method | А | М | А | М | А | М | А | М | А | М | А | М | А | М | А | М |
| 0T | 67.04 | 70.43 | 41.72 | 39.09 | 123.23 | 125.51 | 4879.61 | 4855.96 | 57.98 | 43.66 | 446.77 | 429.19 | 336.34 | 327.01 | 14.34 | 14.46 |
| МТ | 69.22 | 61.40 | 36.05 | 32.99 | 127.60 | 127.80 | 3872.92 | 3428.97 | 39.50 | 28.44 | 496.06 | 449.02 | 300.61 | 273.59 | 11.78 | 10.24 |

Table 1b. Zn, Cu, Mn, Fe concentrations (mg $\rm kg^{-1})$ in rat tissues and rat tapeworm

MT = rats with zinc lactate in feed mixture and tapeworm infection, 0T = control rats with tapeworm infection, A = arithmetic mean, M = me-

dian *weak significance (P < 0.05), **medium significance (P < 0.005), ***strong significance (P < 0.0005)

all computations were done using program STATISTICA Version 10 (Statsoft, USA)

can directly alter the epithelial ion transport, causing increased secretion, decreased ion absorption or both (K o s i k - B o g a c k a e t a l., 2010).

As is evident in Fig 1a, b, tapeworms decreased Zn concentrations in the majority of rat tissues. This phenomenon was significant in the testes and liver tissues in groups with zinc lactate (Fig. 1a). In the control groups, this decrease was significant in the liver, testes, bone, and kidneys (Fig. 1b). This supports the theory regarding the ability of tapeworms to accumulate heavy metals from the host. Hymenolepidiasis is associated with the activation of inflammatory mediators and stimulation of nerve fibres, which significantly affect the function of ion channels in the intestine epithelium of the host (K o s i k - B o g a c k a et al., 2010).

Copper (Cu) is important for proper growth of the body, efficient utilization of iron, proper enzymatic reactions, as well as improved health of connective tissues, hair, and eyes. It is also integral for preventing premature aging and increasing energy production. Apart from these, Cu regulated heart rhythm, balanced thyroid glands, reduced symptoms of arthritis, supported quick wound healing, increased red blood cell formation, and reduced cholesterol (K u c h a r z e w s k i et al., 2003).

Tapeworms significantly decreased Cu concentrations in the testes, liver, and kidneys of rats given overdoses of zinc lactate; surprisingly, Cu concentrations were higher in the spleens of rats with tapeworm infection (Fig. 2a). There were no differences between Cu concentrations in parasitized and unparasitized rats surprisingly, only bone tissues had higher Cu concentrations in rats with tapeworm infection. From these results we can surmise that tapeworm infection has no significant effect on Cu concentrations in host tissues (Fig. 2b). Zn overdosing significantly decreased Cu concentrations in rat spleen (Table 1a) and in the testes of rats with tapeworm infection (Table 1b).

Manganese (Mn) is an essential dietary nutrient and trace element, and at low concentrations it plays an important role in mammalian development, metabolism, and antioxidant defense; however, it becomes neurotoxic at higher concentrations (C h u a, M o r g a n, 1996; M e r c a d a n t e et al., 2016). As we can see in Fig. 3a, b, tapeworms significantly decreased Mn concentrations in the testes, spleen, liver, and

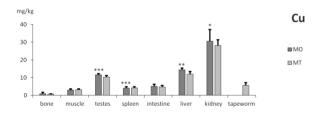


Fig. 2a. Copper concentrations (mg kg-1) in rat tissues and rat tapeworm M0 = rats with zinc lactate in feed mixture, MT = rats with zinc lactate in feed mixture and tapeworm infection statistically significant difference between groups ***P ≥ 0.001 , **P ≥ 0.05 , *P ≥ 0.01

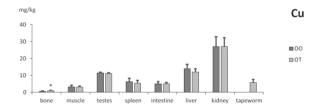
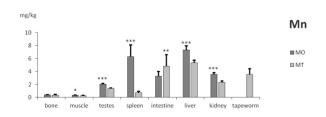


Fig. 2b. Copper concentrations (mg kg–1) in rat tissues and rat tapeworm 00 = control rats, 0T = control rats with tapeworm infection statistically significant difference between groups ***P ≥ 0.001 , **P ≥ 0.05 , *P ≥ 0.01



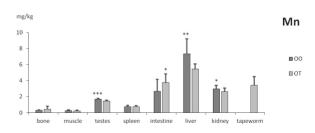


Fig. 3a. Manganese concentrations (mg kg–1) in rat tissues and rat tapeworm M0 = rats with zinc lactate in feed mixture, MT = rats with zinc lactate in feed mixture and tapeworm infection statistically significant difference between groups ***P ≥ 0.001 , **P ≥ 0.05 , *P ≥ 0.01

Fig. 3b. Manganese concentrations (mg kg–1) in rat tissues and rat tapeworm 00 = control rats, 0T = control rats with tapeworm infection statistically significant difference between groups ***P \geq 0.001, **P \geq 0.05, *P \geq 0.01

kidneys of rats given overdoses of zinc lactate; interestingly, rats with tapeworm infection had significantly higher Mn concentrations in their intestinal tissues in both groups (Fig. 3a, b). Rats given a Zn overdose had significantly higher Mn concentrations in their spleen, small intestine, kidneys, testes, bone, and muscle than did rats without a Zn overdose (Table 1a). However, rats given a Zn overdose and infected with tapeworms had significantly lower Mn concentrations in their testes and kidneys than did rats with only tapeworm infection (Table 1b).

Iron is a crucial element for both the pathogen/ parasite and host in the context of a number of infectious diseases (N a v a r r e t e - P e r e a et al., 2016). In vertebrate hosts, parasites can obtain iron from different host sources including erythrocytes, serum hemoglobin, haptoglobin-hemoglobin complexes, hemopexin, transferrin, and lactoferrin (C a s s a t, S k a a r, 2013).

With respect to iron, tapeworms significantly decreased Fe concentrations in the testicular, bone, muscle and liver tissues of rats not given zinc overdose (Fig. 4b). In rats given a zinc overdose, tapeworms significantly decreased Fe concentrations only in the kidneys. Surprisingly, bone tissues of rats infected with tapeworms and given overdoses of zinc lactate (Fig. 4a) had higher Fe concentrations than those of rats without tapeworm infection. Fe concentrations in the livers of rats not given a Zn overdose (Fig. 4b) were significantly lower in rats with tapeworm infection, i.e., tapeworm infection significantly decreased Fe concentrations in the liver. Zn overdosing in our study prevented significant decreases in Fe concentrations in the livers of rats with tapeworm infection (Fig. 4a). However, there were significantly lower Fe concentrations in the kidneys of rats with tapeworms than in those of rats without tapeworms. This indicates that Zn overdosing did not inhibit the tapeworm's ability to decrease Fe in the kidneys (Fig. 4a). When rats were infected with tapeworms (Table 1b), there were no differences in Fe tissue concentrations between rats given a Zn overdose and those not overdosed with Zn. However,

mg/kg Fe 700 600 500 ∎мо 400 m M1 300 200 100 1 E 前子 高子 muscle testes intestine tape

Fig. 4a. Iron concentrations (mg kg-1) in rat tissues and rat tapeworm M0 = rats with zinc lactate in feed mixture, MT = rats with zinc lactate in feed mixture and tapeworm infection

statistically significant difference between groups ***P $\geq 0.001,$ **P

in rats not infected with tapeworms (Table 1a) we found significantly lower Fe concentrations in the bone, muscle, testes, and liver (tissues) of rats given a zinc overdose.

CONCLUSION

Tapeworm presence decreased element concentrations in a majority of rat tissues and tapeworms accumulated more zinc and manganese than did host tissues. Tapeworms can decrease element concentrations in host tissues either through accumulation into their tissues or by increasing intestinal mucus, which decreases ion absorption. Zn overdosing increased Zn concentrations only in the testes, spleen, and tapeworms of rats with tapeworm infection. Moreover, Mn concentrations increased in the spleen, small intestine, kidneys, testes, bone, and muscle of rats given a zinc overdose and not infected with tapeworm. Cu concentrations in rat tissues and the accumulation of Cu, Fe, and Mn by tapeworms were virtually unaffected. Zinc overdosing seems to protect the liver from the absorption of Fe by tapeworms. With the exception of Zn concentrations, element concentrations in tapeworms from hosts given a zinc overdose were similar to those in tapeworms from hosts not given a zinc overdose.

ACKNOWLEDGEMENT

We would like to thank Mr. Brian Kavalir for his proofreading services.

REFERENCES

Brozova A, Jankovska I, Miholova D, Schankova S, Truneckova J, Langrova I, Kudrnacova M, Vadlejch J (2015): Heavy metal concentrations in the small intestine of red fox (*Vulpex* vulpex) with and without *Echinococcus multilocularis* infec-

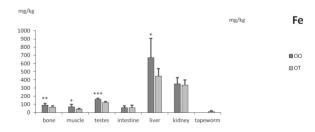


Fig. 4b. Iron concentrations (mg kg-1) in rat tissues and rat tapeworm 00 = control rats, 0T = control rats with tapeworm infection statistically significant difference between groups ***P ≥ 0.001 , **P > 0.05, *P > 0.01

 $\geq 0.05, *P \geq 0.01$

tion. Environmental Science and Pollution Research, 22, 3175–3179. doi: 10.1007/s11356-014-3733-7.

- Burges A, Epelde L, Benito G, Artetxe U, Becerril JM, Garbisu C (2016): Enhancement of ecosystem services during endophyte-assisted aided phytostabilization of metal contaminated mine soil. Science of the Total Environment, 562, 480-492. doi: 10.1016/j.scitotenv.2016.04.080.
- Cadkova Z, Szakova J, Miholova D, Valek P., Pacakova Z, Vadlejch J, Langrova I, Jankovska I (2013): Faecal excretion dynamic during subacute oral exposure to different Pb species in Rattus norvegicus. Biological Trace Element Research, 152, 225–232. doi: 10.1007/s12011-013-9609-8.
- Cassat JE, Skaar EP (2013): Iron in infection and immunity. Cell Host and Microbe, 13, 509–519. doi: 10.1016/j. chom.2013.04.010.
- Chua ACG, Morgan EH (1996): Effects of iron deficiency and iron overload on manganese uptake and deposition in the brain and other organs of the rat. Biological Trace Element Research, 55, 39–54.
- Gil-Jimanez E, Manzano J, Casado E, Ferrer M (2017): The role of density dependence regulation in the misleading effect of the Aznacollar minig spill on the booted eagle fecundity. Science of the Total Environment, 583, 440–446. doi: 10.1016/j.scitotenv.2017.01.098.
- Henriquez-Hernandez LA, Carreton E, Camacho M, Montoya-Alonso JA, Boada LD, Valeron PF, Cordon YF, Almeida-Gonzalez M, Zumbado M, Luzardo OP (2016): Influence of parasitism in dogs on their serum levels of persistent organochlorine compounds and polycyclic aromatic hydrocarbons. Science of the Total Environment, 562, 128–135. doi: 10.1016/j.scitotenv.2016.03.204.
- Jankovska I, Vadlejch J, Szakova J, Miholova D, Kunc P, Knizkova I, Langrova I (2010a): Experimental studies on the lead accumulation in the cestode Moniezia expansa (Cestoda: Anoplocephalidae) and its final host (Ovis aries). Ecotoxicology, 19, 928-932. doi: 10.1007/s10646-010-0474-3.
- Jankovska I, Vadlejch J, Szakova J, Miholova D, Kunc P, Knizkova I, Cadkova Z, Langrova I (2010b): Experimental studies on the cadmium accumulation in the cestode Moniezia expansa (Cestoda: Anoplocephalidae) and its final host (Ovis aries). Experimental Parasitology, 126, 130–134. doi: 10.1016/j.exppara.2010.04.010.
- Jankovska I, Sloup V, Szakova J, Langrova I, Sloup S (2016): How the tapeworm Hymenolepis diminuta affects zinc and cadmium accumulation in a host fed a hyperaccumulating plant (Arabidopsis halleri). Environmental Science and Pollution Research, 23, 19126–19133. doi: 10.1007/ s11356-016-7123-1.
- Jimenez-Diaz I, Artacho-Cordon F, Vela-Soria F, Belhassen H, Arrebola JP, Fernandez MF, Ghali R, Hedhili A, Olea N (2016): Urinary levels of bisphenol A, benzophenones and parabens in Tunisian women: a pilot study. Science of the Total Environment, 562, 81-88. doi: 10.1016/j.scitotenv.2016.03.203.

- Kim JH, Park Y, Kim SK, Moon HB, Park J, Choi K, Kim S (2017): Timing of an accelerated body mass increase in children exposed to lead in early life: a longitudinal study. Science of the Total Environment, 584–585, 72-77. doi: 10.1016/j.scitotenv.2017.01.122.
- Kosik-Bogacka DI, Baranowska-Bosiacka I, Salamatin R (2010): Hymenolepis diminuta: effect of infection on ion transport in colon and blood picture of rats. Experimental Parasitology, 124, 285–294. doi: 10.1016/j.exppara.2009.10.014.
- Kucharzewski M, Braziewicz J, Majewska U, Gozdz S (2003): Copper, zinc, and selenium in whole blood and thyroid tissue of people with various thyroid diseases. Biological Trace Element Research 93, 9–18. doi: 10.1385/BTER:93:1-3:9#page-1.
- Kulma M, Bubova T, Kopecky O, Rettich F (2017): Lavender, eucalyptus, and orange essential oils as repellents against Ixodes ricinus females. Scientia Agriculturae Bohemica, 48, 76-81. doi: 10.1515/sab-2017-0014.
- Kyrianova IA, Vadlejch J, Langrova I (2017): Eimeriosis seasonal dynamics patterns at an organic sheep farm in the Czech Republic. Scientia Agriculturae Bohemica, 48, 70-75. doi: 10.1515/sab-2017-0013.
- Mercadante S, Aielli F, Masedu F, Valenti M, Verna L, Porzio G (2016): Age differences in the last week of life in advanced cancer patients followed at home. Supportive Care in Cancer, 24, 1889–1895. doi: 10.1007/s00520-015-2988-y.
- Navarrete-Perea J, Moguel B, Bobes RJ, Villalobos N, Carrero JC, Sciutto E, Soberon X, Laclette JP (2016): Protein profiles of Taenia solium cysts obtained from skeletal muscles and the central nervous system of pigs: search for tissuespecific proteins. Experimental Parasitology, 172, 23–29. doi: 10.1016/j.exppara.2016.11.006.
- Pavlovic P, Mitrovic M, Dordevic D, Sakan S, Slobodnik J, Liska I, Csanyi B, Jarie S, Kostie O, Pavlovic D, Marinkovic N, Tubic B, Paunovic M (2016): Assessment of the contamination of riparian soil and vegetation by trace metals – a Danube River case study. Science of the Total Environment, 540, 396-409. doi: 10.1016/j.scitotenv.2015.06.125.
- Sloup V, Jankovska I, Langrova I, Stolcova M, Sloup S, Nechybova S, Perinkova P (2016): Changes of some biochemical parameters after the high doses administration of zinc lactate. Scientia Agriculturae Bohemica, 47, 148–153. doi: 10.1515/ sab-2016-0022.
- Sloup V, Jankovska I, Nechybova S, Perinkova P, Langrova I (2017): Zinc in the animal organism: a review. Scientia Agriculturae Bohemica, 48, 13–21. doi: 10.1515/sab-2017-0003.
- Strachel R, Zaborowska M, Wyszkowska J (2016): Deliberations on zinc – a trace mineral or a toxic element? Journal of Elementology, 21, 625-639. doi: 10.5601/jelem.2015.20.3.991.
- Sures B (2004): Environmental parasitology: relevancy of parasites in monitoring environmental pollution. Trends in Parasitology, 20, 170–177. doi: 10.1016/j.pt.2004.01.014.
- Sures B, Franken M, Taraschewski H (2000a): Element concentrations in the archiacanthocephalan Macracanthorhynchus

hirudinaceus compared with those in the porcine definitive host from a slaughterhouse in La Paz, Bolivia. International Journal for Parasitology, 30, 1071–1076. doi: 10.1016/ S0020-7519(00)00094-1.

- Sures B, Jurges G, Taraschewski H (2000b): Accumulation and distribution of lead in the archiacanthocephalan Moniliformis moniliformis from experimentally infected rats. Parasitology, 121, 427-433. doi: 10.1017/S003118209900654X.
- Sures B, Grube K, Taraschewski H (2002): Experimental studies on the lead accumulation in the cestode Hymenolepis diminuta and its final host, Rattus norvegicus. Ecotoxicology, 11, 365–368. doi: 10.1023/A:1020561406624.
- Vymazal J (2017): The use of constructed wetlands for nitrogen removal from agricultural drainage: a review. Scientia Agriculturae Bohemica, 48, 82-91. doi: 10.1515/ sab-2017-0009.

Corresponding Author:

prof. Ing. Ivana J a n k o v s k á , Ph.D., Czech University of Life Sciences Prague, Faculty of Agrobiology, Food and Natural Resources, Department of Zoology and Fisheries, 165 00 Prague 6-Suchdol, Czech Republic, phone: +420 775 986 602, e-mail: jankovska@af.czu.cz